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US DEPARTMENT OF COMMERCE PATENT AND TRADEMARK OFFICE

ATTORNEY DOCKET NUMBER
1416/OP551PCUSTRANSMITTAL LETTER TO THE UNITED STATES
DESIGNATED/ELECTED OFFICE (DO/EO/US)
CONCERNING A FILING UNDER 35 U.S.C. §371U.S. APPLICATION NO.
(if known, see PCT/PTO 1390)
NEW 09/142613International Application No.
PCT/JP97/00804International Filing Date
March 13, 1997Priority Date Claimed
March 13, 1996

Title of Invention

ANTI-PHOSPHORYLATED TAU PROTEIN ANTIBODIES AND METHODS FOR DETECTING ALZHEIMER'S
DISEASE WITH THE USE OF THE SAME

Applicant(s) For DO/EO/US

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Applicant herewith submits to the United States Designated/Elected Office (DO/EO/US) the following items and other information:

1. ☒ This is a **FIRST** submission of items concerning a filing under 35 U.S.C. §371.
2. ☐ This is a **SECOND** or **SUBSEQUENT** submission of items concerning a filing under 35 U.S.C. §371.
3. ☐ This express request to begin national examination procedures (35 U.S.C. §371(f)) at any time rather than delay examination until the expiration of the applicable time limit set in 35 U.S.C. §371(b) and PCT Articles 22 and 39(1).
4. ☒ A proper Demand for International Preliminary Examination was made by the 19th month from the earliest claimed priority date.
5. ☒ A copy of the International Application as filed (35 U.S.C. §371(c)(2))
 - a. ☐ is transmitted herewith (required only if not transmitted by the International Bureau).
 - b. ☒ has been transmitted by the International Bureau. **ATTACHMENT A**
 - c. ☐ is not required, as the application was filed in the United States Receiving Office (RO/US)
6. ☒ A translation of the International Application into English (35 U.S.C. §371(c)(2)). **ATTACHMENT B**
7. ☐ Amendments to the claims of the International Application under PCT Article 19 (35 U.S.C. §371(c)(3)).
 - a. ☐ are transmitted herewith (required only if not transmitted by the International Bureau).
 - b. ☐ have been transmitted by the International Bureau.
 - c. ☐ have not been made; however, the time limit for making such amendments has NOT expired.
 - d. ☐ have not been made and will not be made.
8. ☒ An unexecuted oath or declaration of the inventor(s) (35 U.S.C. §371(c)(4)). **ATTACHMENT C**
9. ☐ A translation of the annexes to the International Preliminary Examination Report under PCT Article 36 (35 U.S.C. §371(c)(5)).

Items 10. to 13. below concern other document(s) or information included:

10. ☒ An Information Disclosure Statement under 37 CFR 1.97 and 1.98. **ATTACHMENT D**
11. ☐ An assignment document for recording. A separate cover sheet in compliance with 37 CFR 3.28 and 3.31 is included.
12. ☐ A **FIRST** preliminary amendment.
☐ A **SECOND** or **SUBSEQUENT** preliminary amendment.
13. ☐ Other items or information:

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U.S. APPLICATION NO. (if known, see 37 CFR 1.5) NEW		INTERNATIONAL APPLICATION NO. PCT/JP97/00804		ATTORNEY'S DOCKET NO. 1416/OP551PCUS	
14. [X] The following fees are submitted BASIC NATIONAL FEE (37 CFR 1.492(a)(1)-(5)): [X] Search Report has been prepared by the EPO or JPO..... \$ 930.00 [] Neither international preliminary examination fee (37 CFR 1.482) nor international search fee (37 CFR 1.445(a)(2)) paid to USPTO..... \$1,070.00 ENTER APPROPRIATE BASIC FEE AMOUNT =				CALCULATIONS	PTO USE ONLY
				\$930.00	
Surcharge of \$130.00 for furnishing the oath or declaration later than [] 20 [] 30 months from the earliest claimed priority date (37 CFR 1.492(e)).				\$	
Claims	Number Filed	Number Extra	Rate		
Total Claims	-20 =		X \$22.00	\$	
Independent Claims	- 3 =		X \$82.00	\$	
Multiple dependent claim(s) (if applicable)			+ \$270.00	\$270.00	
TOTAL OF ABOVE CALCULATIONS =				\$1,200.00	
Reduction by 1/2 for filing by small entity, if applicable. Verified Small Entity Statement must also be filed. (Note 37 CFR 1.9, 1.27, 1.28)				\$	
SUBTOTAL =				\$1,200.00	
Processing fee of \$130.00 for furnishing the English translation later than [] 20 [] 30 months from the earliest claimed priority date (37 CFR 1.492(f)).			+	\$	
TOTAL NATIONAL FEE =				\$1,200.00	
Fee for recording the enclosed assignment (37 CFR 1.21(h)). The assignment must be accompanied by an appropriate cover sheet (37 CFR 3.28, 3.31). \$40 per property +				\$	
TOTAL FEES ENCLOSED =				\$1,200.00	
				Amount to be refunded	\$
				Amount to be charged	\$
a. [X] A check in the amount of \$1,200.00 to cover the above fees is enclosed. A duplicate copy of this form is enclosed. b. [] Please charge my Deposit Account No. 23-0975 in the amount of \$_____ to cover the above fees. A duplicate copy of this sheet is enclosed. c. [] The Commissioner is hereby authorized to charge any additional fees which may be required, or credit any overpayment to Deposit Account No. <u>23-0975</u> . A duplicate copy of this form is enclosed.					
NOTE: Where an appropriate time limit under 37 CFR 1.494 or 1.495 has not been met, a petition to revive (37 CFR 1.137(a) or (b)) must be filed and granted to restore the application to pending status.					
SEND ALL CORRESPONDENCE TO: WENDEROTH, LIND & PONACK, L.L.P. 2033 K St., N.W., Ste. 800 Washington, D.C. 20006			<div style="text-align: center;"> <div style="display: flex; justify-content: center; align-items: center;"> <div style="border-bottom: 1px solid black; width: 150px; margin-bottom: 5px;"></div> <div style="margin-bottom: 5px;">SIGNATURE</div> </div> <div style="display: flex; justify-content: center; align-items: center;"> <div style="border-bottom: 1px solid black; width: 150px; margin-bottom: 5px;"></div> <div style="margin-bottom: 5px;">NAME</div> </div> <div style="display: flex; justify-content: center; align-items: center;"> <div style="border-bottom: 1px solid black; width: 100px; margin-bottom: 5px;"></div> <div style="margin-bottom: 5px;">REGISTRATION NUMBER</div> </div> </div>		
September 11, 1998			[CHECK NO. 29826]		

-1-

DESCRIPTION

ANTI-PHOSPHORYLATED TAU PROTEIN ANTIBODIES AND METHODS
FOR DETECTING ALZHEIMER'S DISEASE WITH THE USE OF THE
SAME

5

Technical Field

The present invention relates to antibodies used for detecting Alzheimer's disease. More specifically, the present invention relates to antibodies to a partial peptide containing phosphorylation sites of phosphorylated tau protein in the paired helical filament, a reagent kit containing the same, and methods for detecting Alzheimer's disease using the antibodies or the kit.

10

Background Art

15

Alzheimer's disease is progressive dementia occurring at the presenile stage (the age of 45 to 65). It causes morbid changes such as degeneration of neurons and atrophy of cerebral cortex due to a decrease in the number of neurons. Pathologically, a number of senile plaques and neurofibrillary degeneration are observed in the brain. So-called senile dementia caused by spontaneous aging in the senium of the age of 65 or

20

older is not substantially different from Alzheimer's disease from the pathological viewpoint and is regarded as senile dementia of Alzheimer type.

5 The number of patients suffering from Alzheimer's disease increases as senile population grows. This disease has thus drawn social attention. There are various hypotheses about the causes of this disease. However, it has not been elucidated yet and it is desired to be clarified soon.

10 The main component of senile plaques that is one of pathological changes caused by Alzheimer's disease is known to be amyloid β protein (Annu. Rev. Neurosci., 12, 463-490 (1989)). Neurofibrillary degeneration that is another pathological change shows accumulation of the
15 paired helical filament in neurons and tau protein is identified as one of its constituents (J. Biochem. 99, 1807-1810 (1986); Proc. Natl. Acad. Sci. USA, 83, 4913-4917 (1986)).

20 Tau protein is composed of a group of protein isoforms that usually produce several bands at the molecular weight of 48 to 65 kD as a result of SDS-polyacrylamide gel electrophoresis and it is known to promote formation of microtubule. Tau protein incorporated in the PHF of the Alzheimer diseased brain
25 was proved to be abnormally phosphorylated as compared with that in the normal brain using polyclonal antibody to PHF (anti-ptau; J. Biochem., 99, 1807-1810 (1986))

and monoclonal antibody to tau protein (tau-1 antibody;
Proc. Natl. Acad. Sci. USA, 83, 4913-4917 (1986)). The
phosphorylation sites of phosphorylated tau protein
incorporated in the PHF were also identified (JP 6-
5 239893 A). Thus, functions of tau protein involved in
Alzheimer's disease has being clarified.

However, it has not been known so far to detect
Alzheimer's disease based on the phosphorylation site of
phosphorylated tau protein in the PHF. Although methods
10 for detecting Alzheimer's disease using various
antibodies have been proposed, a clinically effective
new detection method is still desired.

Disclosure of the Invention

The present inventors paid attention to the
15 phosphorylation sites of phosphorylated tau protein in
the PHF and found that antibodies obtained by using as
an immunogen a partial peptide comprising a
phosphorylated sites are useful for detecting
Alzheimer's disease, thereby completing the present
20 invention.

The present invention provides antibodies obtained
by using as an immunogen a partial peptide comprising a
phosphorylation sites of phosphorylated tau protein in
the paired helical filament.

25 Preferred embodiments of this invention provides:

the antibody as described above, wherein the phosphorylation site of phosphorylated tau protein is one or more amino acid residues selected from serine at position 198, serine at position 199, serine at position 202, threonine at position 205, threonine at position 231, serine at position 235, serine at position 262, serine at position 396, threonine at position 403, serine at position 404, serine at position 409, serine at position 412, serine at position 413, and serine at position 422 of an amino acid sequence of SEQ ID NO: 1;

the antibody as described above, wherein the phosphorylation site of phosphorylated tau protein is one or more amino acid residues selected from serine at position 199, serine at position 202, threonine at position 205, threonine at position 231, serine at position 235, serine at position 262, serine at position 396, serine at position 404, serine at position 412, serine at position 413, and serine at position 422 of the amino acid sequence of SEQ ID NO: 1;

the antibody as described above, wherein the partial peptide comprising the phosphorylation site comprises an amino acid residue at the phosphorylated site and plural amino acid residues before and/or after sites of the phosphorylation site; and

the antibody as described above, wherein the partial peptide comprising the phosphorylation site has an amino acid sequence of any one of SEQ ID NO: 2 to SEQ

ID NO: 16.

Another embodiment of this invention provides a reagent kit used for detecting Alzheimer's disease comprising at least any one of the above-described
5 antibodies.

Yet another embodiment of this invention provides methods for detecting Alzheimer's disease comprises examining reactivity between any one of the above-described antibodies and a sample from an individual
10 suspected of Alzheimer's disease.

The present invention will be described in detail below.

In the present invention, an example of tau protein is that having a primary structure consisting of 352 to
15 441 amino acid residues as described in Goedert et al., Neuron 3, 519-526 (1989). For example, when tau protein having the primary structure represented by the amino acid sequence shown in SEQ ID NO: 1 in the sequence listing is used, one or more amino acid residues to be
20 phosphorylated are selected from serine at position 198, serine at position 199, serine at position 202, threonine at position 205, threonine at position 231, serine at position 235, serine at position 262, serine at position 396, threonine at position 403, serine at
25 position 404, serine at position 409, serine at position 412, serine at position 413, and serine at position 422 of the same sequence (J. Biol. Chem., 270, 823-829

(1995) and Neurosci. Lett., 189, 167-170 (1995)).

It is preferable to use a partial peptide of tau protein comprising one or more phosphorylated amino acid residues selected from serine at position 199, serine at
5 position 202, threonine at position 205, threonine at position 231, serine at position 235, serine at position 262, serine at position 396, serine at position 404, serine at position 412, serine at position 413 and serine at position 422 of the above-described sequence.

10 A peptide comprising the amino acid residue(s) at the above-described phosphorylated site(s) and plural amino acid residues before and/or after sites of the phosphorylation site(s) is preferably used as the partial peptide of tau protein used in the present
15 invention. In particular, such a peptide comprises preferably 1 to 7 amino acid residues, more preferably 3 to 5 amino acid residues, at either before or after site or both sites of the amino acid residue at the phosphorylation site. Among these partial peptides, the
20 peptide having the amino acid sequence described in any one of SEQ ID NO: 2 to 16 in the sequence listing is most preferably used.

The antibody of the present invention can be obtained by immunizing an animal with the partial
25 peptide comprising the phosphorylation site of the phosphorylated tau protein of the present invention as an immunogen and preparing serum from the animal. In

this occasion, it is preferable to use as an immunogen a carrier protein to which the above-described peptide having introduced at its amino terminus or carboxyl terminus an amino acid residue with a reactive functional group, such as cysteine, lysine, glutamic acid, or aspartic acid, is bound as a hapten.

Among the above-described peptides, the partial peptides represented by SEQ ID NO: 3, 13, 15, and 16 are synthesized by the solid phase peptide synthesis using a phenyl group as a protective group for the phosphorylation site as described in Tetrahedron Lett., 32, 7083-7086 (1991).

The phosphorylated peptides represented by the other sequence than those represented by the above-described sequence identification number, which have an aromatic amino acid, a sulfur-containing amino acid, or a heterocyclic amino acid, are synthesized by the solid phase peptide synthesis using a cyclohexyl group as a protective group for the phosphorylation site as described in Peptide Chemistry 1993, 109-112 (1994) or by the solid phase peptide synthesis using a benzyl group as a protective group for the phosphorylation site as described in Chem. Lett., 1099-1112 (1994).

In the preparation of the antibody, the partial peptide synthesized as described above is bound to a carrier protein such as bovine serum albumin (BSA), thyroglobulin, or keyhole limpet hemocyanin. The

binding can be performed easily using an appropriate
condensing agent, such as maleimide, glutaraldehyde, or
carbodiimide. The thus-obtained peptide bound to a
carrier protein is used to immunize an animal. The
5 immunization of an animal can be performed in the same
method as conventional methods used for antibody
production. More specifically, a solution comprising
the peptide bound to a carrier protein is mixed with
adjuvant if necessary, and subcutaneously or
10 intraperitoneally administered to animals usually used
for antibody production, such as mice, rats, rabbits,
guinea pigs, or sheep. Additional immunization is
performed every two to three weeks after the first
immunization to produce antiserum with a high antibody
15 titer. Blood is collected from the immunized animal to
prepare serum. In the present invention, the thus-
obtained antiserum can be used as it is without
purification. Alternatively, serum may be heat-treated
to inactivate complements and subjected to salting out
20 with ammonium sulfate and ion exchange chromatography to
purify immunoglobulin-containing fractions.
Furthermore, the antibody may be purified using a
peptide column having a specific partial peptide
immobilized to obtain an antibody specifically
25 recognizing the above-described phosphorylation site or
its vicinity.

A monoclonal antibody recognizing a specific

epitope can be obtained by collecting antibody-producing cells from the immunized animal, subjecting the collected cells to cell fusion with cultured cells such as a myeloma cell line derived from the same animal to
5 prepare hybridoma, and preparing immunoglobulin fractions from the culture medium of the hybridoma by conventional methods.

Alzheimer's disease can be detected by immunologically reacting the antibody obtained by the
10 present invention with a sample derived from an individual suspected of Alzheimer's disease by per se known conventional methods, detecting an antigen-antibody reaction product, and examining reactivity between the sample and the antibody. The antibody
15 obtained as described above can be used to prepare a reagent kit for detecting Alzheimer's disease by the above-described method comprising the immunoreaction and the detection step. Such a kit comprises the constituents usually used in the immunoreaction-based
20 kit. More specifically, the reagent kit of the present invention comprises at least the antibody of the present invention and, as optional ingredients, a solution for diluting a sample, a washing solution, a labeled antibody or a labeled antigen, chromogen, and a peptide
25 for positive control.

Alzheimer's disease can be detected using the antibody or the reagent kit of the present invention for

example as follows.

A sample is first obtained from an individual suspected of Alzheimer's disease and reacted with the antibody obtained as described above. The sample may be
5 tissues from cerebral cortex or the like and body fluid such as cerebrospinal fluid or blood. When a tissue sample is subjected to the detection method of the present invention, about 0.1 mg of the sample is required. When cerebrospinal fluid or blood is used as
10 a sample, about 0.5 to 0.01 ml of the sample is required.

Once the above-described sample is obtained, the sample is homogenized in physiological saline and centrifuged. The resulting supernatant is fractionated
15 to remove contaminating immunoglobulin and examined for reactivity to the above-obtained antibody as an index.

The thus-obtained fraction is electrophoresed. The antibody as obtained above is added thereto to perform immunoblotting. In this occasion, the antibody can be
20 detected by labeling it with the label used conventionally. Alternatively, the antibody may be detected by reacting it with a secondary antibody that is reactive with the antibody.

If a sample from an individual suspected of
25 Alzheimer's disease is examined for reactivity with the antibody and the reactivity increases compared with a control derived from an individual without Alzheimer's

disease, the individual is regarded to be a patient of Alzheimer's disease. If the reactivity decreases compared with a control from an individual with Alzheimer's disease, the individual is regarded to be free from Alzheimer disease. In this way, Alzheimer's disease can be detected by the present invention.

In the present invention, unlike an antibody obtained using as an immunogen phosphorylated tau protein as a whole or the PHF, the antibody highly specific to each phosphorylation site can be obtained. This antibody is useful for detection specific to the phosphorylation site of phosphorylated tau protein without determining antigen-recognizing specificity. Furthermore, various antibodies site-specific to each phosphorylation site can be efficiently obtained. The thus-obtained various antibodies are examined for reactivity with a sample from an individual with Alzheimer's disease to select an antibody most suitable for detection of Alzheimer's disease easily.

Brief Description of the Drawings

Figure 1 is the dot blot showing specificity of the antibodies obtained by immunization with a partial peptide containing a phosphorylation site of phosphorylated tau protein.

Figure 2 is photographs of electrophoresis

(immunoblotting) showing reactivity of the TS fraction
(the fraction obtained by removing IgG from the
supernatant of human cerebral cortex suspension)
obtained in Example with the antibodies used in the
5 present invention.

Figure 3 is photographs of electrophoresis
(immunoblotting) showing reactivity of the SDS
precipitation fraction obtained in Example with the
antibodies used in the present invention.

10 Figure 4 is photographs of electrophoresis
(immunoblotting) showing reactivity of the SDS
precipitation fraction obtained in Example with the
antibodies used in the present invention.

Figure 5 shows a calibration curve in the
15 competitive RIA obtained in Example.

Figure 6 shows the results of measuring the
concentrations of phosphorylated tau protein in the
cerebrospinal fluid from patients with Alzheimer's
disease and patients with no dementia obtained in
20 Example.

Best Mode for Carrying out the Invention

The present invention will be described below in
more detail with reference to Examples, but is not
construed to be limited thereto.

Production Example 1

Preparation of partial peptide having amino acid
sequence described in SEQ ID NO: 3 in the sequence
listing (This peptide and the antibodies against this
5 peptide are hereinafter sometimes referred to as "PS202"
and "anti-PS202", respectively.)

A peptide shown by H-Lys-Ser-Ser-Pro-Gly-
Ser(H₂PO₃)-Pro-Gly-Thr-Pro-Gly-Ser-Arg-NH₂ (SEQ ID NO: 3)
was produced by the following method. Symbols
10 hereinafter used respectively have the following
meanings: MBHA resin: p-methylbenzhydrylamine resin;
Boc: t-butyloxycarbonyl group; Bzl: benzyl group; Ph:
phenyl group; Tos: p-toluenesulfonyl group; and Z(2-Cl):
2-chlorobenzoyloxycarbonyl group.

15 (a) Production of H-Lys[Z(2-Cl)]-Ser(Bzl)-Ser(Bzl)-Pro-G
ly-Ser[PO(OPh)₂]-Pro-Gly-Thr(BzL)-Pro-Gly-Ser(Bzl)-Arg(T
os)-MBHA resin

0.94 g of MBHA resin (amine content: 0.64 mmol/g
resin) was set in the Bio Search 9500 Model Automatic
20 Peptide Synthesizer. Boc-Arg(Tos)-OH, Boc-Ser(Bzl)-OH,
Boc-Gly-OH, Boc-Pro-OH, Boc-Thr(Bzl)-OH, Boc-Gly-OH,
Boc-Pro-OH, Boc-Ser[PO(OPh)₂]-OH, Boc-Gly-OH, Boc-Pro-
OH, Boc-Ser(Bzl)-OH, Boc-Ser(Bzl)-OH, and Boc-Lys[Z(2-
Cl)]-OH were supplied thereto and were then coupled in
25 this order using diisopropylcarbodiimide as a condensing
agent to obtain 2.38 g of the above-described side
chain-protected peptide-MBHA resin.

(b) Treatment with hydrogen fluoride

1.34 g of the side chain-protected peptide-MBHA resin obtained in (a) was collected and set in a hydrogen fluoride reaction device manufactured by the Peptide Institute, Inc.. The resin was reacted with 13 ml of hydrogen fluoride in the presence of 1.5 ml anisole under ice-cooling for 1 hour. After the completion of the reaction, hydrogen fluoride was distilled off under reduced pressure. The residue was washed with ethyl acetate and extracted with 150 ml of 2 M acetic acid to obtain 350 mg of a crude peptide having a protected phosphate group and shown by H-Lys-Ser-Ser-Pro-Gly-Ser[PO(OPh)₂]-Pro-Gly-Thr-Pro-Gly-Ser-Arg-NH₂.

This product was dissolved in 20 ml of 30% acetic acid and the resulting solution was applied to a Sephadex G-25 column (internal diameter: 5 cm and length: 109 cm). Elution was performed using the same solvent to collect the fraction containing the desired product. The thus-obtained fraction was then dissolved in a small amount of distilled water and purified by HPLC using a reverse phase column (internal diameter: 2 cm and length: 25 cm) packed with ODS (octadecylsilane)-bound silica. Elution was performed by a linear gradient of 5 to 65% acetonitrile in 0.1% trifluoroacetic acid. The yield of the purified product was 110 mg. The structure of this substance was confirmed by FAB mass spectrometry; measured [M + H]⁺;

1445, calculated ($C_{61}H_{92}N_{18}O_{21}P_1 + H$); 1445.

(c) Hydrogenolysis

90 mg of the phosphate group-protected peptide
obtained in (b) and 80 mg of platinum oxide (catalyst)
5 were mixed with 1 ml of acetic acid and the mixture was
stirred at room temperature for 12 hours under hydrogen
atmosphere of 5 to 6 pressure. After the catalyst was
filtered off, the filtrate and washings were collected
and lyophilized. The resulting product was purified by
10 preparatory HPLC to obtain 55 mg of a final product,
phosphorylated peptide shown by H-Lys-Ser-Ser-Pro-Gly-
Ser[H_2PO_3]-Pro-Gly-Thr-Pro-Gly-Ser-Arg-NH₂. The
structure of this substance was confirmed by FAB mass
spectrometry; measured $[M + H]^+$; 1294, calculated
15 ($C_{49}H_{85}N_{18}O_{21}P_1 + H$); 1294.

Production Examples 2 to 4

Partial peptides having amino acid sequences
described in SEQ ID NO: 13, NO: 15, and NO: 16 in the
sequence listing were obtained in the same manner as
20 described in Production Example 1 (These peptides are
hereinafter sometimes referred to as "PS413", "PS412",
and "PS412, 413" and the antibodies to these peptides
are referred to as "anti-PS413", "anti-PS412", and
"anti-PS412, 413", respectively.)

Preparation of partial peptide having amino acid
sequence described in SEQ ID NO: 2 in the sequence
listing (This peptide and the antibodies to this peptide
are hereinafter sometimes referred to as "PS199" and
5 "anti-PS199", respectively.)

A peptide shown by H-Lys-Ser-Gly-Tyr-Ser-
Ser(H₂PO₃)-Pro-Gly-Ser-Pro-Gly-Thr-NH₂ (SEQ ID NO: 2) was
produced by the following method. Symbols used
hereinafter respectively have the following meanings:
10 MBHA resin: p-methylbenzhydrylamine resin; Boc: t-
butyloxycarbonyl group; Bzl: benzyl group; cHex:
cyclohexane group; Z(2-Br): 2-bromobenzyloxycarbonyl
group; and Z(2-Cl): 2-chlorobenzyloxycarbonyl group.
(a) Production of H-Lys[Z(2-Cl)]-Ser(Bzl)-Gly-Tyr[Z(2-Br
15)]-Ser(Bzl)-Ser[PO(OcHex)₂]-Pro-Gly-Ser(BzL)-Pro-Gly-Thr
(Bzl)-MBHA resin

131 mg of MBHA resin (amine content: 0.76 mmol/g
resin) was set in the Bio Search 9500 Model Automatic
Peptide Synthesizer. Boc-Thr(Bzl)-OH, Boc-Gly-OH, Boc-
20 Pro-OH, Boc-Ser(Bzl)-OH, Boc-Gly-OH, Boc-Pro-OH, Boc-
Ser[PO(OcHex)₂]-OH, Boc-Ser(Bzl)-OH, Tyr[Z(2-Br)]-OH,
Boc-Gly-OH, Boc-Ser(Bzl)-OH, and Boc-Lys[Z(2-Cl)]-OH
were supplied thereto and were then coupled in this
order using diisopropylcarbodiimide as a condensing
25 agent to obtain 307 mg of the above-described side
chain-protected peptide-MBHA resin.

(b) Treatment with trifluoromethanesulfonic acid

150 mg of the side chain-protected peptide-MBHA resin obtained in (a) was collected. 10 ml of trifluoroacetic acid containing 1 M methanesulfonic acid and thioanisole, and 0.05 ml of m-cresol were added thereto and the resulting mixture was allowed to react for 4 hours under ice-cooling. After the completion of the reaction, 200 ml of ice-cooled diethyl ether was added thereto to precipitate a peptide. The whole content was collected by filtration with a glass filter and washed with cold diethyl ether. The residue was extracted with 200 ml of 2 M acetic acid to obtain 53 mg of a crude peptide shown by H-Lys-Ser-Gly-Tyr-Ser-Ser[H₂PO₃]-Pro-Gly-Ser-Pro-Gly-Thr-NH₂.

(c) Purification of the peptide

This product was dissolved in 6 ml of distilled water and purified by HPLC using a reverse phase column (internal diameter: 2 cm and length: 25 cm) packed with ODS (octadecylsilane)-bound silica. Elution was performed by a linear gradient of 5 to 35% acetonitrile in 0.1% trifluoroacetic acid. The yield of the purified product was 29 mg. The structure of this substance was confirmed by FAB mass spectrometry; measured [M + H]⁺; 1204, calculated (C₄₇H₇₅N₁₄O₂₁P₁ + H); 1204.

Production Example 6

Preparation of partial peptide having amino acid sequence described in SEQ ID NO: 6 in the sequence

listing (This peptide and the antibodies to this peptide are hereinafter sometimes referred to as "PT231" and "anti-PT231", respectively.)

A peptide shown by H-Cys-Val-Ala-Val-Val-Arg-
5 Thr(H₂PO₃)-Pro-Pro-Lys-Ser-Pro-Ser-Ser-OH (SEQ ID NO: 6)
was produced by the following method. Symbols used
hereinafter respectively have the following meanings:
Bzl resin: benzyl alcohol resin; Boc: t-butyloxycarbonyl
group; Bzl: benzyl group; MBzl: 4-methoxybenzyl group;
10 Mts: methylenesulfonyl group; cHex: cyclohexyl group;
and Z(2-Cl): 2-chlorobenzylloxycarbonyl group.

(a) Production of H-Cys(MBzl)-Val-Ala-Val-Val-Arg(Mts)-
Thr[PO(OcHex)₂]-Pro-Pro-Lys[Z(2-Cl)]-Ser(BzL)-Pro-Ser(Bz
1) -Ser(Bzl)-Bzl resin

15 71 mg of Boc-Ser(Bzl)-Bzl resin (amine content:
0.70 mmol/g resin) was set in the Bio Search 9500 Model
Automatic Peptide Synthesizer. Boc-Ser(Bzl)-OH, Boc-
Pro-OH, Boc-Ser(Bzl)-OH, Boc-Lys[Z(2-Cl)]-OH, Boc-Pro-
OH, Boc-Pro-OH, Boc-Thr[PO(OcHex)₂]-OH, Boc-Arg(Mts)-OH,
20 Boc-Val-OH, Boc-Val-OH, Boc-Ala-OH, Boc-Val-OH, and Boc-
Cys(MBzl)-OH were supplied thereto and were then coupled
in this order using diisopropylcarbodiimide as a
condensing agent to obtain 62 mg of the above-described
side chain-protected peptide-Bzl resin.

25 (b) Treatment with trifluoromethanesulfonic acid

To 62 mg of the side chain-protected peptide-Bzl
resin obtained in (a) were added 0.9 ml of

trifluoromethanesulfonic acid, 1.2 ml of thioanisole,
6.6 ml of trifluoroacetic acid, 0.9 ml of m-cresol, and
0.4 ml of ethanedithiol. The mixture was allowed to
react for 5 minutes under ice-cooling and then at room
5 temperature for 3 hours. After the completion of the
reaction, 200 ml of ice-cooled diethyl ether was added
thereto to precipitate a peptide. The whole content was
collected by filtration with a glass filter and washed
with cold diethyl ether. The residue was extracted with
10 170 ml of 2 M acetic acid to obtain 21 mg of a crude
peptide shown by H-Cys-Val-Ala-Val-Val-Arg-Thr(H₂PO₃)-
Pro-Pro-Lys-Ser-Pro-Ser-Ser-OH.

(c) Purification of the peptide

This product was dissolved in 10 ml of 30% acetic
15 acid and applied to a Sephadex G-25 column (internal
diameter: 5 cm and length: 107 cm). Elution was
performed using the same solvent to collect the
fractions containing the desired product. The yield of
this purified product was 12 mg.

20 The thus-obtained product was dissolved in 5 ml of
30% acetic acid and purified by HPLC using a reverse
phase column (internal diameter: 2 cm and length: 25 cm)
packed with ODS (octadecylsilane)-bound silica. Elution
was performed with 13% acetonitrile in 0.1%
25 trifluoroacetic acid. The yield of the purified product
was 7 mg. The structure of this substance was confirmed
by FAB mass spectrometry; measured [M + H]⁺; 1509,

calculated ($C_{61}H_{107}N_{18}O_{22}P_1S_1 + H$): 1508.

Production Example 7

Preparation of partial peptide having amino acid
sequence described in SEQ ID NO: 11 in the sequence
listing (This peptide and the antibodies to this peptide
are hereinafter sometimes referred to as "PS396" and
"anti-PS396", respectively.)

A peptide shown by H-Cys-Glu-Ile-Val-Tyr-Lys-
Ser(H_2PO_3)-Pro-Val-Val-Ser-Gly-NH₂ (SEQ ID NO: 11) was
produced by the following method. Symbols used
hereinafter respectively have the following meanings:
MBHA resin: p-methylbenzhydrylamine resin; Boc: t-
butyloxycarbonyl group; Bzl: benzyl group; MBzl, 4-
methoxybenzyl group; cHex: cyclohexyl group; Z(2-Br): 2-
bromobenzyloxycarbonyl group; and Z(2-Cl): 2-
chlorobenzyloxycarbonyl group.

(a) Production of H-Cys(MBzl)-Glu(OBzl)-Ile-Val-Tyr[Z(2-
Br)]-Lys[Z(2-Cl)]-Ser[PO(OcHex)₂]-Pro-Val-Val-Ser(Bzl)-Gly-MBHA resin

131 mg of MBHA resin (amine content: 0.76 mmol/g resin) was set in the Bio Search 9500 Model Automatic Peptide Synthesizer. Boc-Gly-OH, Boc-Ser(Bzl)-OH, Boc-Val-OH, Boc-Val-OH, Boc-Pro-OH, Boc-Ser[PO(OcHex)₂]-OH, Boc-Lys[Z(2-Cl)]-OH, Tyr[Z(2-Br)]-OH, Boc-Val-OH, Boc-Ile-OH, Boc-Glu(OBzl)-OH, and Boc-Cys(MBzl)-OH were supplied thereto and were then coupled in this order

using diisopropylcarbodiimide as a condensing agent to obtain 376 mg of the above-described side chain-protected peptide-MBHA resin.

(b) Treatment with trifluoromethanesulfonic acid

5 To 188 mg of the side chain-protected peptide-MBHA resin obtained in (a) were added 10 ml of trifluoroacetic acid containing 1 M methanesulfonic acid and thioanisole, and 0.05 ml of m-cresol. The mixture was allowed to react for 4 hours under ice-cooling.

10 After the completion of the reaction, 200 ml of ice-cooled diethyl ether was added thereto to precipitate a peptide. The whole content was collected by filtration with a glass filter and washed with cold diethyl ether, and extracted with 200 ml of 2 M acetic acid to obtain

15 87 mg of a crude peptide shown by H-Cys-Glu-Ile-Val-Tyr-Lys-Ser(H₂PO₃)-Pro-Val-Val-Ser-Gly-NH₂.

(c) Purification of the peptide

 This product was dissolved in 9 ml of 30% acetic acid and purified by HPLC using a reverse phase column

20 (internal diameter: 2 cm and length: 25 cm) packed with ODS (octadecylsilane)-bound silica. Elution was performed with 16% acetonitrile in 0.1% trifluoroacetic acid. The yield of the purified product was 45 mg. The structure of this substance was confirmed by FAB mass

25 spectrometry; measured [M + H]⁺; 1360, calculated (C₅₇H₉₅N₁₄O₂₀P₁S₁ + H); 1360.

Production Examples 8 to 10

Partial peptides having amino acid sequence described in SEQ ID NO: 4, NO: 12, and NO: 14 in the sequence listing were obtained in the same manner as in Production Examples 5, 6, and 7 (These peptides are hereinafter referred to as "PT205", "PS404", and "PS422", respectively, and antibodies to these peptides are referred to as "anti-PT205", "anti-PS404", and "anti-PS422", respectively.)

Production Example 11

Preparation of partial peptide having amino acid sequence described in SEQ ID NO: 7 in the sequence listing (This peptide and the antibodies to this peptide are hereinafter sometimes referred to as "PS235" and "anti-PS235", respectively.)

A peptide shown by H-Cys-Arg-Thr-Pro-Pro-Lys-Ser(H₂PO₃)-Pro-Ser-Ser-Ala-Lys-OH (SEQ ID NO: 7) was produced by the following method. Symbols used hereinafter respectively have the following meanings:

Alko resin: p-alkoxybenzyl alcohol resin; Boc: t-butyloxycarbonyl group; tBu: t-butyl group; Bzl: benzyl group; Fmoc: 9-fluorenylmethoxycarbonyl group; Trt: trityl group; Pmc: pentamethylchroman-6-sulfonyl group.

(a) Production of H-Cys(Trt)-Arg(Pmc)-Thr(tBu)-Pro-Pro-Lys(Boc)-Ser[PO(OH)(OBzl)]-Pro-Ser(tBu)-Ser(tBu)-Ala-Lys(Boc)-Alko resin

385 mg of Fmoc-Lys(Boc)-Alko resin (amino acid content: 0.65 mmol/g resin) was set in the Applied Bio Systems 431 Model Automatic Peptide Synthesizer. Fmoc-Ala-OH, Fmoc-Ser(tBu)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Pro-OH, Fmoc-Ser[PO(OH)(OBzl)]-OH, Fmoc-Lys(Boc)-OH, Fmoc-Pro-OH, Fmoc-Pro-OH, Fmoc-Thr(tBu)-OH, and Fmoc-Arg(Pmc)-OH were supplied thereto and were then coupled in this order using HBTU [2-(1H-benzotriazole-1-yl)-1,1,3,3-tetramethyluronium hexafluorophosphate] as a condensing agent to obtain 716 mg of a side chain-protected peptide-Alko resin intermediate. Fmoc-Cys(Trt)-OH was condensed with 358 mg of this intermediate to obtain 395 mg of the above-described side chain-protected peptide-Alko resin.

(b) Treatment with trifluoroacetic acid

To 196 mg of the side chain-protected peptide-Alko resin obtained in (a) was added a mixture containing 8.25 ml of trifluoroacetic acid, 0.5 ml of purified water, 0.5 ml of thioanisole, 0.75 ml of phenol, and 0.25 ml of ethanedithiol. The resulting mixture was allowed to react for 1.5 hour at room temperature. After the completion of the reaction, 200 ml of ice-cooled diethyl ether was added thereto to precipitate a peptide. The whole content was collected by filtration with a glass filter, washed with cold diethyl ether, and extracted with 80 ml of 2 M acetic acid to obtain 82 mg of a crude peptide shown by H-Cys-Arg-Thr-Pro-Pro-Lys-

Ser(H₂PO₃)-Pro-Ser-Ser-Ala-Lys-OH.

(c) Purification of the peptide

This product was dissolved in 7 ml of 0.1% trifluoroacetic acid and purified by HPLC using a reverse phase column (internal diameter: 2 cm and length: 25 cm) packed with ODS (octadecylsilane)-bound silica. Elution was performed with 7% acetonitrile in 0.1% trifluoroacetic acid. The yield of the purified product was 62 mg. The structure of this substance was confirmed by FAB mass spectrometry; measured [M + H]⁺; 1339, calculated (C₅₂H₉₂N₁₇O₂₀P₁S₁ + H); 1339.

Production Examples 12 to 15

Partial peptides having amino acid sequence described in SEQ ID NO: 5, NO: 8, NO: 9, and NO: 10 in the sequence listing were obtained in the same manner as in Production Example 11 (These peptides are hereinafter sometimes referred to as "PS199,202", "ratPS235", "PT231,PS235", and "PS262", respectively, and the antibodies to these peptides are referred to as "anti-PS199,202", "anti-ratPS235", "anti-PT231,PS235", and "anti-PS262", respectively.)

Production Example 16

Preparation of partial peptide having amino acid sequence described in SEQ ID NO: 17 in the sequence listing (This peptide and the antibodies to this peptide

are hereinafter sometimes referred to as "Tau-C" and "anti-Tau-C", respectively.)

A peptide shown by H-Ser-Pro-Gln-Leu-Ala-Thr-Leu-Ala-Asp-Glu-Val-Ser-Ala-Ser-Leu-Ala-Lys-OH (SEQ ID NO: 17) was produced by the following method. Symbols used hereinafter respectively have the following meanings: Alko resin: p-alkoxybenzyl alcohol resin; Boc: t-butyloxycarbonyl group; tBu: t-butyl group; Bzl: benzyl group; Fmoc: 9-fluorenylmethoxycarbonyl group; and Trt: trityl group.

(a) Production of H-Ser(tBu)-Pro-Gln(Trt)-Leu-Ala-Thr(tBu)-Leu-Ala-Asp(OtBu)-Glu(OtBu)-Val-Ser(tBu)-Ala-Ser-Leu-Ala-Lys(Boc)-Alko resin

284 mg of Alko resin (amine content: 0.88 mmol/g resin) was set in the ABI A431 Model Automatic Peptide Synthesizer. Fmoc-Lys(Boc)-OH was bound to the resin using dimethylaminopyridine and diisopropylcarbodiimide as condensing agents. Then, Fmoc-Ala-OH, Fmoc-Leu-OH, Fmoc-Ser(tBu)-OH, Fmoc-Ala-OH, Fmoc-Ser(tBu)-OH, Fmoc-Val-OH, Fmoc-Glu(OtBu)-OH, Fmoc-Asp(OtBu)-OH, Fmoc-Ala-OH, Fmoc-Leu-OH, Fmoc-Thr(tBu)-OH, Fmoc-Ala-OH, Fmoc-Leu-OH, Fmoc-Gln(Trt)-OH, Fmoc-Pro-OH, and Fmoc-Ser(tBu)-OH were supplied thereto and were then coupled in this order using HBTU [2-(1H-benzotriazole-1-yl)-1,1,3,3-tetramethyluronium hexafluorophosphate] as a condensing agent to obtain 905 mg of the above-described side chain-protected peptide-Alko resin.

(b) Treatment with trifluoroacetic acid

To 543 mg of the side chain-protected peptide-Alko resin obtained in (a) were added 9.5 ml of trifluoroacetic acid, 0.25 ml of ethanedithiol, and 0.5 ml of distilled water. The mixture was allowed to react for 5 minutes under ice-cooling and then at room temperature for 1.5 hour. After the completion of the reaction, 200 ml of ice-cooled diethyl ether was added thereto to precipitate a peptide. The whole content was collected by filtration with a glass filter, washed with cold diethyl ether, and extracted with 100 ml of 2 M acetic acid to obtain 250 mg of a crude peptide shown by H-Ser-Pro-Gln-Leu-Ala-Thr-Leu-Ala-Asp-Glu-Val-Ser-Ala-Ser-Leu-Ala-Lys-OH.

(c) Purification of the peptide

This product was dissolved in 20 ml of 30% acetic acid and applied to a Sephadex G-25 column (internal diameter: 5 cm and length: 107 cm). Elution was performed using the same solvent to collect the fractions containing the desired product. The yield of this purified product was 122 mg. The structure of this substance was confirmed by FAB mass spectrometry; measured $[M + H]^+$; 1702, calculated $(C_{73}H_{125}N_{19}O_{27}S_2 + H)$; 1701.

Production Example 17

Preparation of partial peptide having amino acid

sequence described in SEQ ID NO: 18 in the sequence listing (This peptide and the antibodies to this peptide are hereinafter sometimes referred to as "Tau-N" and "anti-Tau-N", respectively.)

5 A peptide shown by H-Ala-Glu-Pro-Arg-Gln-Glu-Glu-Phe-Glu-Val-Met-Glu-Cys-NH₂ (SEQ ID NO: 18) was produced by the following method. Symbols used hereinafter respectively have the following meanings: Fmoc-NH-SAL resin: 4-(2',4'-dimethoxyphenyl-Fmoc-aminoethyl)phenoxy
10 resin; Boc: t-butyloxy-carbonyl group; tBu: t-butyl group; Bzl: benzyl group; Fmoc: 9-fluorenylmethoxycarbonyl group; Trt: trityl group; and Pmc: pentamethylchroman-6-sulfonyl group.
(a) Production of H-Ala-Glu(OtBu)-Pro-Arg(Pmc)-Gln(Trt)-
15 Glu(OtBu)-Phe-Glu(OtBu)-Val-Met-Glu(OtBu)-Cys(Trt)-NH-SAL resin

532 mg of Fmoc-NH-SAL resin (amine content: 0.47 mmol/g resin) was set in the ABI A431 Model Automatic Peptide Synthesizer. Fmoc-Cys(Trt)-OH, Fmoc-Glu(OtBu)-
20 OH, Fmoc-Met-OH, Fmoc-Val-OH, Fmoc-Glu(OtBu)-OH, Fmoc-Phe-OH, Fmoc-Glu(OtBu)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Arg(Pmc)-OH, Fmoc-Pro-OH, Fmoc-Glu(OtBu)-OH, and Fmoc-Ala-OH were supplied thereto and were then coupled in this order using HBTU [2-(1H-benzotriazole-1-yl)-1,1,3,
25 3-tetramethyluronium hexafluorophosphate] as a condensing agent to obtain 1122 mg of the above-described side chain-protected peptide-NH-SAL resin.

(b) Treatment with trifluoroacetic acid

To 673 mg of the side chain-protected peptide-NH-SAL resin obtained in (a) were added 0.75 ml of phenol, 0.5 ml of thioanisole, 8.25 ml of trifluoroacetic acid, 0.25 ml of ethanedithiol, and 0.5 ml of distilled water. The mixture was allowed to react for 5 minutes under ice-cooling and then at room temperature for 1.5 hour. After the completion of the reaction, 200 ml of ice-cooled diethyl ether was added thereto to precipitate a peptide. The whole content was collected by filtration with a glass filter, washed with cold diethyl ether, and extracted with 50 ml of 2 M acetic acid and 250 ml of distilled water to obtain 182 mg of a crude peptide shown by H-Ala-Glu-Pro-Arg-Gln-Glu-Glu-Phe-Glu-Val-Met-Glu-Cys-NH₂.

(c) Purification of the peptide

This product was dissolved in 20 ml of 30% acetic acid and applied to a Sephadex G-25 column (internal diameter: 5 cm and length: 107 cm). Elution was performed using the same solvent to collect the fractions containing the desired product. The yield of this purified product was 136 mg.

This product was dissolved in 20 ml of 20% acetonitrile and purified by HPLC using a reverse phase column (internal diameter: 2 cm and length: 25 cm) packed with ODS (octadecylsilane)-bound silica. Elution was performed with 22% acetonitrile in 0.1%

trifluoroacetic acid. The yield of the purified product was 96 mg. The structure of this substance was confirmed by FAB mass spectrometry; measured $[M + H]^+$; 1467, calculated $(C_{61}H_{95}N_{17}O_{21}S_2 + H)$; 1467.

5

Example 1

Preparation of antibodies

The partial peptides obtained in Production Examples 1 to 17 were each bound to an equivalent weight of keyhole limpet hemocyanin to serve as an immunogen.

10 0.2 mg of each of the immunogens was dissolved in 0.3 ml of physiological saline and emulsified with an equivalent volume of Freund's adjuvant. Rabbits were immunized with the resulting emulsion every three weeks. The thus-obtained antisera were purified by applying

15 onto a Affigel 15 column (Bio-Rad) to which the respective antigenic peptide was bound and eluted using Immunopure gentle Ag/Ab buffer system (Pierce). Thus, the above-described antibodies that specifically recognize the respective phosphorylation sites were

20 obtained.

Specificity of the antibodies was confirmed by dot blotting.

More specifically, Immobilon P-membrane (Millipore) was dotted with 18 pmol of each peptide dissolved in a

25 70% DMSO solution in line with each other, allowed to adsorb them, and dried. This membrane was immersed in

TBS (20 mM Tris-HCl (pH 7.5), 150 mM NaCl) containing 5% skim milk for 1 hour to prevent non-specifically binding of the antibodies to be added in the following step to the membrane. The membrane was then washed with TBS for 5 minutes three times, immersed in TBS containing the desired antibody (first antibody), placed between parafilm, and allowed to react at 4°C for 14 hours in a humidified box to bind the first antibodies to the respective antigen peptides on the membrane. The membrane was then washed with TBS containing 0.05% Tween 20 (TBST) for 5 minutes three times. The following procedure until color development was performed using ProtoBlot Western Blot AP System (Promega).

Anti-rabbit IgG antibody to which alkaline phosphatase was covalently bound (second antibody) was diluted 5000-fold with TBST. The membrane was immersed in this diluted solution at 4°C for 2 hours to bind alkaline phosphatase to the antigen-first antibody binding product on the membrane through the second antibody. The membrane was washed with TBST for 5 minutes three times, then with TBS for 5 minutes twice. The membrane was immersed in a reaction mixture (100 mM Tris-HCl (pH 9.5), 100 mM NaCl, 5 mM MgCl₂) supplemented with 0.165 mg/ml of 5-bromo-4-chloro-3-indolyl phosphate (BCIP) and 0.33 mg/ml of nitro blue tetrazolium (NBT) and the existence of alkaline phosphatase on the membrane was detected by development of purple color.

The reaction was terminated by immersing the membrane in water.

The first antibodies used in this Example were all rabbit antisera as they were. The same results were
5 obtained using IgG obtained by purifying the antisera by affinity chromatography using a peptide column. The dilution of each antiserum was 1000-fold for anti-PS199, 250-fold for anti-PS202, 500-fold for anti-PT205, 250-fold for anti-PT231, 250-fold for anti-PS235, 25-fold
10 for anti-rat PS235 (anti-rPS235), 500-fold for anti-PS262, 1000-fold for anti-PS396, 500-fold for anti-PS404, 500-fold for anti-PS413, and 500-fold for anti-PS422. The control peptides used were non-phosphorylated peptides including the amino acid sequence shown by SEQ
15 ID NO: 19 (a peptide shown by K1, the 226th to the 240th amino acid residues, in SEQ ID NO: 1 in Fig. 1), the amino acid sequence shown by SEQ ID NO: 20 (a peptide shown by K2, the 191st to the 224th amino acid residues, in SEQ ID NO: 1 in Fig. 1), the amino acid sequence
20 shown by SEQ ID NO: 21 (a peptide shown by AK-K3, the 384th to the 438th amino acid residues, in SEQ ID NO: 1 in Fig. 1), and the amino acid sequence shown by SEQ ID NO: 22 (a peptide shown by S262, the 257th to the 267th amino acid residues having a cysteine residue at the N-
25 terminus, in SEQ ID NO: 1 in Fig. 1). These peptides were produced in the same manner in (a) and (b) in Production Example 1.

The results of dot blotting are shown in Fig. 1. The abscissa shows the peptide adsorbed by dotting it on the membrane and the ordinate shows the antibody. It shows that the respective antibodies obtained as described above are specifically bound to the corresponding phosphorylation sites.

Example 2

Study on reactivity of antibody with samples

(1) Preparation of human brain extract

Human brain extracts were prepared from 8 cases of normal human brains and 19 cases of human brains of patients with Alzheimer's disease. The following procedure was all performed at 4°C.

1 g was sampled from chilled specimen of postmortem human cerebral cortex, cut into pieces with a razor in 3 ml of a TSinh solution [50 mM Tris-HCl (pH 7.6), 0.15 M NaCl, 0.5 mM DIFP (diisopropylfluorophosphate), 1 µg/ml antipain, 0.5 mM PMSF (phenylmethanesulfonyl fluoride), 1 mg/ml of TLCK (tricyl-lysine-chloromethyl ketone), 1 µg/ml of leupeptin, 0.1 g/ml of pepstatin], sonicated, and made into a suspension with a homogenizer. The resulting suspension was centrifuged at 80,000 rpm for 15 minutes to obtain a supernatant. Human IgG in this supernatant was removed with Protein G-Sepharose 4 Fast Flow (Pharmacia) and the resulting fraction was designated as TS fraction. The precipitate was

sonicated in 2 ml of the above-described TSinh solution and made into a suspension with a homogenizer. The resulting suspension was washed by centrifugation at 80,000 rpm for 15 minutes and the thus-obtained precipitate was sonicated in 2 ml of TSinh containing 1% Triton X-100 and made into a suspension with a homogenizer. The homogenized product was centrifuged at 80,000 rpm for 15 minutes to obtain a supernatant (TX fraction). The precipitate was sonicated in 2 ml of TSinh containing 1% Triton X-100, made into a suspension with a homogenizer and washed by centrifugation at 80,000 rpm for 15 minutes. The resulting precipitate was sonicated in 2 ml of TSinh containing 2% SDS and homogenized to obtain a suspension. The suspension was centrifuged at 80,000 rpm for 15 minutes to obtain a supernatant (SDS supernatant fraction). The resulting precipitate was sonicated in 2 ml of TSinh containing 2% SDS, made into a suspension with a homogenizer, and washed by centrifugation at 80,000 for 15 minutes. The thus-obtained precipitate was sonicated in 2 ml of TSinh containing 2% SDS and made into a suspension (SDS precipitate fraction) with a homogenizer.

(2) Immunoblotting using phosphorylation site-specific antibodies

Laemmli's sample treatment solution (Nature 227, 680-685 (1970)) was added to each fraction obtained above. The mixture was heated at 95°C for 5 minutes and

subjected to SDS polyacrylamide gel electrophoresis. The resulting electrophoretic patterns were transferred to Immobilon P-membrane (Millipore). After blocking with TBS containing 5% skim milk for 2 hours, the
5 membrane was reacted with the phosphorylation site-specific antibody obtained in Example 1 as a first antibody for 14 hours. The reaction mixture was washed with Tween 20 and TBS and treated with ProtoBlot AP (Promega) as a second antibody to develop color of the
10 antibody-antigen reaction product on the membrane.

The first antibody was all rabbit IgG and dilution of the first antibody was shown by the following numerals after "x" under the names of the respective antibodies in Figs. 2 to 4 with respect to Examples.
15 Almost all of the first antibodies were previously purified by affinity chromatography using an antigen peptide column. However, PS262 and PS422 were used in the form of antisera and distinguished from others by indicating "S" under the name of antibodies in these
20 figures.

Tau-N and Tau-C are peptides obtained in Production Examples 16 and 17 and correspond to the 2nd to the 12th and the 422nd to the 438th amino acid residues, respectively, of human tau protein represented by SEQ ID
25 NO: 1 in the sequence listing, and used as controls showing the existence of tau protein.

A positive control used was whole brain extract of

8-day-old juvenile rat. Juvenile tau protein is known to be highly phosphorylated tau protein which is similar to that in the PHF (J. Biol. Chem. 268, 25712-25717 (1993)). Specifically, 0.75 g of 8-day-old juvenile rat
5 brain was homogenized in 1.5 ml of medium containing 10 mM Tris-HCl (pH 7.4), 50 mM NaCl, 50 mM NaF, 1 mM EDTA, 1 mM EGTA, 50 mM β -glycerophosphoric acid, 0.1 mM Na_3VO_4 , 1 mM PMSF, 1 $\mu\text{g/ml}$ antipain, 1 $\mu\text{g/ml}$ leupeptin, 0.1 $\mu\text{g/ml}$ pepstatin, and 3 mM benzamidine. The
10 homogenized product was centrifuged at 25,000 rpm for 20 minutes and a supernatant was recovered to serve as a positive control.

The results are shown in Figs. 2, 3 and 4. Figure 2 shows the result of immunoblotting of the TS fraction; Figures 3 and 4 show the result of immunoblotting of the
15 SDS precipitate fraction. In these figures, " A " in lane AD/N stands for the result of brain extract from an Alzheimer's disease patient; " N ", for the result of normal brain extract; " M ", for molecular weight
20 markers; the next numerals for the molecular weight of the markers in terms of kD. Rat P8 lane stands for the immunoblotting result of the positive control and the band indicated by an arrow is phosphorylated tau protein band.

25 Any antibody did not react with the normal brain extract, but did react with the brain extract from an Alzheimer's disease patient. This indicates that the

method of the present invention enables detecting
Alzheimer's disease. The TS fraction contains easily
soluble tau protein, whereas the SDS precipitation
fraction contains less soluble tau protein. The method
5 of the present invention enables detecting not only
phosphorylated tau protein in tissues but also that in
cerebrospinal fluid or blood thought to contain easily
soluble tau protein. In other words, cerebrospinal
fluid and blood can also be used as a sample as well as
10 tissues.

Example 3

Studies on reactivity of anti-human phosphorylated tau protein antibodies with a sample by radioimmunoassay (RIA)

15 (1) Preparation of ^{125}I -labeled antigen

(a) Labeling by Bolton-Hunter method

Peptides having a lysine residue at the amino
terminus as shown in Production Examples 1 to 5 (SEQ ID
NO: 3, 13, 15, 16, and 2 in the sequence listing) were
20 labeled with ^{125}I in the following manner.

A solution of ^{125}I -Bolton-Hunter reagent in benzene
(18.5 MBq, 500 μCi ; Du Pont, NEX-120) was placed in a
reaction test tube and benzene was vaporized in a stream
of nitrogen. 20 μg of the peptide dissolved in 50 μl of
25 500 mM sodium chloride-160 mM borate buffer (pH 8.5) was
added to the test tube and reacted for 2 hours under

cooling with occasional stirring. 10 μ l of 10% potassium iodide solution and 700 μ l of 0.1% trifluoroacetic acid were added to the reaction mixture and the resulting mixture was purified by HPLC using a
5 reverse phase column (internal diameter: 4 mm and length: 25 cm) packed with ODS (octadecylsilane)-bound silica. Elution was performed by a linear gradient of acetonitrile in 0.1% trifluoroacetic acid up to 60% at a rate of 1%/min. The flow rate of the eluate was 1
10 ml/min. The eluate was fractionated per 0.5 minute. A part of the fraction considered as a peak monitored by a radioactivity detector was measured with a scintillation counter to confirm a desired fraction. Phosphate buffer (Dulbecco's PBS(-), pH 7.3) containing 2% bovine serum
15 albumin (BSA), 0.05% sodium azide, and 0.05% Tween 20 was added to the fraction. The resulting solution was divided into vials in small amounts containing 1,000,000 to 4,000,000 count/min (cpm) per vial and lyophilized.

(b) Labeling by chrolamin-T method

20 Peptide having a tyrosine residue at the amino terminus in place of a cysteine residue in the peptides as shown in Production Examples 6 to 14 (SEQ ID NO: 6, 11, 4, 12, 14, 7, 5, 8, and 9 in the sequence listing) were synthesized by the Fmoc method as used in
25 Production Example 11 and the resulting peptides were labeled with 125 I in the following manner.

10 μ g of the peptide was dissolved in 50 μ l of 0.2

M phosphate buffer (pH 7.3) ,18.5 MBq of Na¹²⁵I (500 µCi; Du Pont, NEZ-033H) was added thereto, and the mixture was stirred. 3 µg of chrolamin-T (1 mg/ml in 0.2 M phosphate buffer; dissolved just prior to addition) was added thereto. The mixture was stirred and reacted at room temperature for 5 minutes. 17.5 µg of ascorbic acid (0.7 mg/ml 0.2 M phosphate buffer; dissolved just prior to addition) was then added thereto, the mixture was stirred, and the reaction was performed at room temperature for 1 minute. 10 µl of a 10% potassium iodide solution and 700 µl of 0.1% trifluoroacetic acid were added thereto. The resulting mixture was purified by reverse phase HPLC in the same manner as in (a) and fractionated. The desired fraction was divided into vials in small amounts and lyophilized.

(2) Competitive RIA

"Anti-PS262" among the antibodies prepared in Production Example 18 is exemplified below.

¹²⁵I-YPS-262 obtained in (1) was dissolved in RIA buffer (Dulbecco's PBS(-) containing 0.1% BSA, 0.05% sodium azide, and 0.05% Tween 20) and a 100 µl portion (about 10,000 cpm) was added to an assay tube. As a standard, 100 µl of RIA buffer containing peptide YPS262 whose amino acids were determined was also added within the range from 0 to 10000 fmol/ml. After 300 µl of the RIA buffer was added, 100 µl of "anti-PS262" diluted 10,000-fold with the RIA buffer was further added and

the mixture was stirred. The mixture was incubated overnight at 4°C. 100 μ l of normal rabbit serum diluted 128-fold with the RIA buffer, 100 μ l of anti-rabbit IgG goat serum diluted 32-fold, 200 μ l of Dulbecco's PBS(-) containing 8% polyethylene glycol (PEG6000) and 0.2% cellulose powder (Avicel, Registered Trademark) were added thereto. The mixture was stirred and incubated at 4°C for 30 minutes then centrifuged at 4°C at 3000 rpm for 20 minutes. The resulting supernatant was removed by suction and the radioactivity of the precipitation was measured with a scintillation counter.

A calibration curve was prepared by plotting the concentration of the standard substance (fmol/ml) as abscissa and the ratio of the count at each concentration of the standard substance to the count at 0 fmol/ml of its concentration as ordinate. Figure 5 shows this calibration curve. Figure 5 indicates that the peptide can be measured 30 fmol/ml at minimum with good accuracy by the method of the present invention.

Furthermore, a derivative S262 of the peptide shown by SEQ ID NO: 10 in the sequence listing, in which cysteine is not added and serine is not phosphorylated, was synthesized by the Fmoc method in the same manner as in Production Example 11 and was used to evaluate the specificity of the antibody anti-PS262. The count did not decrease even when S262 was added to 2000 fmol/ml, indicating that the antibody of the present invention

specifically recognized PS262 containing phosphorylated serine.

(3) Measuring phosphorylated tau protein in cerebrospinal fluids from patients

5 The concentrations of phosphorylated tau protein in cerebrospinal fluids from 8 patients with Alzheimer's disease (AD) were measured using the method and the calibration curve in (2). The concentrations of phosphorylated tau protein in cerebrospinal fluids from
10 7 non-dementia patients were also measured as a control (CTL). The results are shown in Fig. 6. As shown in Fig. 6, phosphorylated tau protein was not detected in any of CTL, while 45 to 75 fmol/ml of phosphorylated tau protein was detected in AD.

15 Industrial Applicability

 The present invention provides antibodies that specifically recognize phosphorylated tau protein using as an immunogen a partial peptide containing a phosphorylation site of phosphorylated tau protein in
20 the PHF. Phosphorylated tau protein in brain extracts and tissue sections can be detected using the antibody obtained as above. Furthermore, the method of measuring phosphorylated tau protein using this antibody of the present invention enables simply measuring
25 phosphorylated tau protein in body fluids and is useful for detecting Alzheimer's disease.

SEQUENCE LISTING

(1) GENERAL INFORMATION

(i) APPLICANT: Koichi, Ishiguro
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Tsuneko, Uchida
Kazutomo Imahori

(ii) TITLE OF INVENTION: ANTI-PHOSPHORYLATED TAU PROTEIN ANTIBODY AND
METHOD FOR DETECTING ALZHEIMER'S DISEASE WITH THE USE OF THE SAME

(iii) NUMBER OF SEQUENCES: 22

(iv) COMPUTER READABLE FORM:

(A) MEDIUM TYPE: Floppy disk
(B) COMPUTER: IBM PC compatible
(C) OPERATING SYSTEM: PC-DOS/MS-DOS
(D) SOFTWARE: PatentIn Release #1.0, Version #1.30 (EPO)

(vi) CURRENT APPLICATION DATA

(A) APPLICATION NUMBER:
(B) FILING DATE:
(C) CLASSIFICATION:

(vi) PRIOR APPLICATION DATA

(A) APPLICATION NUMBER: JP 8/56090
(B) FILING DATE: 13-MAR-1996

(2) INFORMATION FOR SEQ ID NO: 1:

(i) SEQUENCE CHARACTERISTICS:

(A) SEQUENCE LENGTH: 441 amino acids
(B) SEQUENCE TYPE: amino acid
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 1:

Met	Ala	Glu	Pro	Arg	Gln	Glu	Phe	Glu	Val	Met	Glu	Asp	His	Ala	Gly
1				5					10					15	
Gln	Asp	Thr	Tyr	Gly	Leu	Gly	Asp	Arg	Lys	Asp	Gln	Gly	Gly	Tyr	Thr
			20					25					30		
Met	His	Gln	Glu	Gly	Asp	Thr	Asp	Ala	Gly	Leu	Lys	Glu	Ser	Pro	Leu

35	40	45
Gln Thr Pro Thr Glu Asp Gly Ser Glu Glu Pro Gly Ser Glu Thr Ser		
50	55	60
Asp Ala Lys Ser Thr Pro Thr Ala Glu Asp Val Thr Ala Pro Leu Val		
65	70	75
Asp Glu Gly Ala Pro Gly Lys Gln Ala Ala Ala Gln Pro His Thr Glu		
85	90	95
Ile Pro Glu Gly Thr Thr Ala Glu Glu Ala Gly Ile Gly Asp Thr Pro		
100	105	110
Ser Leu Glu Asp Glu Ala Ala Gly His Val Thr Gln Ala Arg Met Val		
115	120	125
Ser Lys Ser Lys Asp Gly Thr Gly Ser Asp Asp Lys Lys Ala Lys Gly		
130	135	140
Ala Asp Gly Lys Thr Lys Ile Ala Thr Pro Arg Gly Ala Ala Pro Pro		
145	150	155
Gly Gln Lys Gly Gln Ala Asn Ala Thr Arg Ile Pro Ala Lys Thr Pro		
165	170	175
Pro Ala Pro Lys Thr Pro Pro Ser Ser Gly Glu Pro Pro Lys Ser Gly		
180	185	190
Asp Arg Ser Gly Tyr Ser Ser Pro Gly Ser Pro Gly Thr Pro Gly Ser		
195	200	205
Arg Ser Arg Thr Pro Ser Leu Pro Thr Pro Pro Thr Arg Glu Pro Lys		
210	215	220
Lys Val Ala Val Val Arg Thr Pro Pro Lys Ser Pro Ser Ser Ala Lys		
225	230	235
Ser Arg Leu Gln Thr Ala Pro Val Pro Met Pro Asp Leu Lys Asn Val		
245	250	255
Lys Ser Lys Ile Gly Ser Thr Glu Asn Leu Lys His Gln Pro Gly Gly		
260	265	270
Gly Lys Val Gln Ile Ile Asn Lys Lys Leu Asp Leu Ser Asn Val Gln		
275	280	285
Ser Lys Cys Gly Ser Lys Asp Asn Ile Lys His Val Pro Gly Gly Gly		
290	295	300
Ser Val Gln Ile Val Tyr Lys Pro Val Asp Leu Ser Lys Val Thr Ser		
305	310	315
Lys Cys Gly Ser Leu Gly Asn Ile His His Lys Pro Gly Gly Gly Gln		
325	330	335
Val Glu Val Lys Ser Glu Lys Leu Asp Phe Lys Asp Arg Val Gln Ser		
340	345	350
Lys Ile Gly Ser Leu Asp Asn Ile Thr His Val Pro Gly Gly Gly Asn		

66T44:6T34463

	355		360		365	
Lys	Lys	Ile	Glu	Thr	His	Lys
	370		375		380	
Lys	Thr	Asp	His	Gly	Ala	Glu
385		390		395		400
Gly	Asp	Thr	Ser	Pro	Arg	His
	405		410		415	
Ile	Asp	Met	Val	Asp	Ser	Pro
	420		425		430	
Ser	Ala	Ser	Leu	Ala	Lys	Gln
	435		440			

(2) INFORMATION FOR SEQ ID NO: 2:

(i) SEQUENCE CHARACTERISTICS:

- (A) SEQUENCE LENGTH: 12 amino acids
- (B) SEQUENCE TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(ix) FEATURE:

- (A) NAME/KEY :
- (B) LOCATION : 6
- (D) OTHER INFORMATION:Xaa=phosphoserine

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 2:

Lys	Ser	Gly	Tyr	Ser	Xaa	Pro	Gly	Ser	Pro	Gly	Thr
1				5						10	

(2) INFORMATION FOR SEQ ID NO: 3:

(i) SEQUENCE CHARACTERISTICS:

- (A) SEQUENCE LENGTH: 13 amino acids
- (B) SEQUENCE TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(ix) FEATURE:

- (A) NAME/KEY :
- (B) LOCATION : 6
- (D) OTHER INFORMATION:Xaa=phosphoserine

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 3:

Lys	Ser	Ser	Pro	Gly	Xaa	Pro	Gly	Thr	Pro	Gly	Ser	Arg
1				5						10		

(2) INFORMATION FOR SEQ ID NO: 4:

(i) SEQUENCE CHARACTERISTICS:

- (A) SEQUENCE LENGTH: 12 amino acids
- (B) SEQUENCE TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(ix) FEATURE:

- (A) NAME/KEY :
- (B) LOCATION : 7
- (D) OTHER INFORMATION:Xaa=phosphothreonine

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 4:

Cys Pro Gly Ser Pro Gly Xaa Pro Gly Ser Arg Ser
1 5 10

(2) INFORMATION FOR SEQ ID NO: 5:

(i) SEQUENCE CHARACTERISTICS:

- (A) SEQUENCE LENGTH: 13 amino acids
- (B) SEQUENCE TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(ix) FEATURE:

- (A) NAME/KEY :
- (B) LOCATION : 6
- (D) OTHER INFORMATION:Xaa=phosphoserine

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 5:

Lys Ser Xaa Pro Gly Xaa Pro Gly Thr Pro Gly Ser Arg
1 5 10

(2) INFORMATION FOR SEQ ID NO: 6:

(i) SEQUENCE CHARACTERISTICS:

- (A) SEQUENCE LENGTH: 14 amino acids
- (B) SEQUENCE TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(ix) FEATURE:

- (A) NAME/KEY :
- (B) LOCATION : 7
- (D) OTHER INFORMATION:Xaa=phosphothreonine

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 6:

Cys Val Ala Val Val Arg Xaa Pro Pro Lys Ser Pro Ser Ser

1 5 10

(2) INFORMATION FOR SEQ ID NO: 7:

(i) SEQUENCE CHARACTERISTICS:

(A) SEQUENCE LENGTH: 12 amino acids

(B) SEQUENCE TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(ix) FEATURE:

(A) NAME/KEY :

(B) LOCATION : 7

(D) OTHER INFORMATION:Xaa=phosphoserine

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 7:

Cys Arg Thr Pro Pro Lys Xaa Pro Ser Ser Ala Lys

1 5 10

(2) INFORMATION FOR SEQ ID NO: 8:

(i) SEQUENCE CHARACTERISTICS:

(A) SEQUENCE LENGTH: 12 amino acids

(B) SEQUENCE TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(ix) FEATURE:

(A) NAME/KEY :

(B) LOCATION : 7

(D) OTHER INFORMATION:Xaa=phosphoserine

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 8:

Cys Arg Thr Pro Pro Lys Xaa Pro Ser Ala Ser Lys

1 5 10

(2) INFORMATION FOR SEQ ID NO: 9:

(i) SEQUENCE CHARACTERISTICS:

(A) SEQUENCE LENGTH: 12 amino acids

(B) SEQUENCE TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(ix) FEATURE:

(A) NAME/KEY :

(B) LOCATION : 3

(D) OTHER INFORMATION:Xaa=phosphothreonine

(ix) FEATURE:

(A) NAME/KEY :

(B) LOCATION : 7

(D) OTHER INFORMATION:Xaa=phosphoserine

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 9:

Cys Arg Xaa Pro Pro Lys Xaa Pro Ser Ser Ala Lys
1 5 10

(2) INFORMATION FOR SEQ ID NO: 10:

(i) SEQUENCE CHARACTERISTICS:

(A) SEQUENCE LENGTH: 12 amino acids

(B) SEQUENCE TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(ix) FEATURE:

(A) NAME/KEY :

(B) LOCATION : 7

(D) OTHER INFORMATION:Xaa=phosphoserine

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 10:

Cys Lys Ser Lys Ile Gly Xaa Thr Glu Asn Leu Lys
1 5 10

(2) INFORMATION FOR SEQ ID NO: 11:

(i) SEQUENCE CHARACTERISTICS:

(A) SEQUENCE LENGTH: 12 amino acids

(B) SEQUENCE TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(ix) FEATURE:

(A) NAME/KEY :

(B) LOCATION : 7

(D) OTHER INFORMATION:Xaa=phosphoserine

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 11:

Cys Glu Ile Val Tyr Lys Xaa Pro Val Val Ser Gly
1 5 10

(2) INFORMATION FOR SEQ ID NO: 12:

(i) SEQUENCE CHARACTERISTICS:

(A) SEQUENCE LENGTH: 12 amino acids

(B) SEQUENCE TYPE: amino acid

(D) TOPOLOGY: linear
(ii) MOLECULE TYPE: peptide
(ix) FEATURE:
 (A) NAME/KEY :
 (B) LOCATION : 7
 (D) OTHER INFORMATION:Xaa=phosphoserine
(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 12:
Cys Val Ser Gly Asp Thr Xaa Pro Arg His Leu Ser
1 5 10

(2) INFORMATION FOR SEQ ID NO: 13:
 (i) SEQUENCE CHARACTERISTICS:
 (A) SEQUENCE LENGTH: 12 amino acids
 (B) SEQUENCE TYPE: amino acid
 (D) TOPOLOGY: linear
 (ii) MOLECULE TYPE: peptide
 (ix) FEATURE:
 (A) NAME/KEY :
 (B) LOCATION : 7
 (D) OTHER INFORMATION:Xaa=phosphoserine
 (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 13:
Lys Leu Ser Asn Val Ser Xaa Thr Gly Ser Ile Asp
1 5 10

(2) INFORMATION FOR SEQ ID NO: 14:
 (i) SEQUENCE CHARACTERISTICS:
 (A) SEQUENCE LENGTH: 12 amino acids
 (B) SEQUENCE TYPE: amino acid
 (D) TOPOLOGY: linear
 (ii) MOLECULE TYPE: peptide
 (ix) FEATURE:
 (A) NAME/KEY :
 (B) LOCATION : 7
 (D) OTHER INFORMATION:Xaa=phosphoserine
 (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 14:
Cys Ile Asp Met Val Asp Xaa Pro Gln Leu Ala Thr
1 5 10

(2) INFORMATION FOR SEQ ID NO: 15:
 (i) SEQUENCE CHARACTERISTICS:

(A) SEQUENCE LENGTH: 12 amino acids
(B) SEQUENCE TYPE: amino acid
(D) TOPOLOGY: linear
(ii) MOLECULE TYPE: peptide
(ix) FEATURE:
 (A) NAME/KEY :
 (B) LOCATION : 6
 (D) OTHER INFORMATION:Xaa=phosphoserine
(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 15:
Lys Leu Ser Asn Val Xaa Ser Thr Gly Ser Ile Asp
1 5 10

(2) INFORMATION FOR SEQ ID NO: 16:
 (i) SEQUENCE CHARACTERISTICS:
 (A) SEQUENCE LENGTH: 12 amino acids
 (B) SEQUENCE TYPE: amino acid
 (D) TOPOLOGY: linear
 (ii) MOLECULE TYPE: peptide
 (ix) FEATURE:
 (A) NAME/KEY :
 (B) LOCATION : 6
 (D) OTHER INFORMATION:Xaa=phosphoserine
 (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 16:
Lys Leu Ser Asn Val Xaa Xaa Thr Gly Ser Ile Asp
1 5 10

(2) INFORMATION FOR SEQ ID NO: 17:
 (i) SEQUENCE CHARACTERISTICS:
 (A) SEQUENCE LENGTH: 17 amino acids
 (B) SEQUENCE TYPE: amino acid
 (D) TOPOLOGY: linear
 (ii) MOLECULE TYPE: peptide
 (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 17:
Ser Pro Gln Leu Ala Thr Leu Ala Asp Glu Val Ser Ala Ser Leu Ala Lys
1 5 10 15

(2) INFORMATION FOR SEQ ID NO: 18:

(i) SEQUENCE CHARACTERISTICS:

- (A) SEQUENCE LENGTH: 12 amino acids
- (B) SEQUENCE TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 18:

Ala Glu Pro Arg Gln Glu Phe Glu Val Met Glu Cys
1 5 10

(2) INFORMATION FOR SEQ ID NO: 19:

(i) SEQUENCE CHARACTERISTICS:

- (A) SEQUENCE LENGTH: 15 amino acids
- (B) SEQUENCE TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 19:

Val Ala Val Val Arg Thr Pro Pro Lys Ser Pro Ser Ser Ala Lys
1 5 10 15

(2) INFORMATION FOR SEQ ID NO: 20:

(i) SEQUENCE CHARACTERISTICS:

- (A) SEQUENCE LENGTH: 34 amino acids
- (B) SEQUENCE TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 20:

Ser Gly Asp Arg Ser Gly Tyr Ser Ser Pro Gly Ser Pro Gly Thr Pro
1 5 10 15
Gly Ser Arg Ser Arg Thr Pro Ser Leu Pro Thr Pro Pro Thr Arg Glu
 20 25 30
Pro Lys

(2) INFORMATION FOR SEQ ID NO: 21:

(i) SEQUENCE CHARACTERISTICS:

- (A) SEQUENCE LENGTH: 55 amino acids
- (B) SEQUENCE TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 21:

-50-

Ala Lys Thr Asp His Gly Ala Glu Ile Val Tyr Lys Ser Pro Val Val
1 5 10 15
Ser Gly Asp Thr Ser Pro Arg His Leu Ser Asn Val Ser Ser Thr Gly
20 25 30
Ser Ile Asp Met Val Asp Ser Pro Gln Leu Ala Thr Leu Ala Asp Glu
35 40 45
Val Ser Ala Ser Leu Ala Lys
50 55

(2) INFORMATION FOR SEQ ID NO: 22:

(i) SEQUENCE CHARACTERISTICS:

(A) SEQUENCE LENGTH: 12 amino acids

(B) SEQUENCE TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 22:

Cys Lys Ser Lys Ile Gly Ser Thr Glu Asn Leu Lys
1 5 10

What is claimed is:

1. Antibodies obtained by using as an immunogen a partial peptide comprising phosphorylation sites of phosphorylated tau protein in a paired helical filament.

5 2. Antibodies defined in claim 1, recognizing the phosphorylation site(s) of phosphorylated tau protein which site(s) are one or more amino acid residues selected from serine at position 198, serine at position 199, serine at position 202, threonine at position 205,
10 threonine at position 231, serine at position 235, serine at position 262, serine at position 396, threonine at position 403, serine at position 404, serine at position 409, serine at position 412, serine at position 413, and serine at position 422 of an amino
15 acid sequence of SEQ ID NO: 1.

 3. Antibodies defined in claim 1, recognizing the phosphorylation site(s) of phosphorylated tau protein which site(s) are one or more amino acid residues selected from serine at position 199, serine at position
20 202, threonine at position 205, threonine at position 231, serine at position 235, serine at position 262, serine at position 396, serine at position 404, serine at position 412, serine at position 413, and serine at position 422 of an amino acid sequence of SEQ ID NO: 1.

25 4. Antibodies defined in any one of claims 1 to 3, which recognize the partial peptide comprising the

phosphorylation site(s) which peptide comprises an amino acid residue at the phosphorylation site(s) and plural amino acid residues before and/or after the phosphorylation site.

5 5. Antibodies defined in any one of claims 1 to 4, which recognize the partial peptide comprising the phosphorylation site, which peptide has an amino acid sequence of any one of SEQ ID NO: 2 to SEQ ID NO: 16.

10 6. A reagent kit used for detecting Alzheimer's disease, comprising at least the antibodies as defined in any one of claims 1 to 5.

15 7. Methods for detecting Alzheimer's disease, which are characterized by examining reactivity of the antibodies, as defined in any one of claims 1 to 5, with a sample from an individual suspected of Alzheimer's disease.

Abstract

5 An antibody is prepared by using a partial peptide
containing the phosphorylated site of a phosphorylated
tau protein in the paired helical filament as an
immunogen. Then the reactivity of the antibody thus
obtained with samples obtained from individuals with a
suspicion of Alzheimer's disease is examined to thereby
detect the disease.

FIG. 1

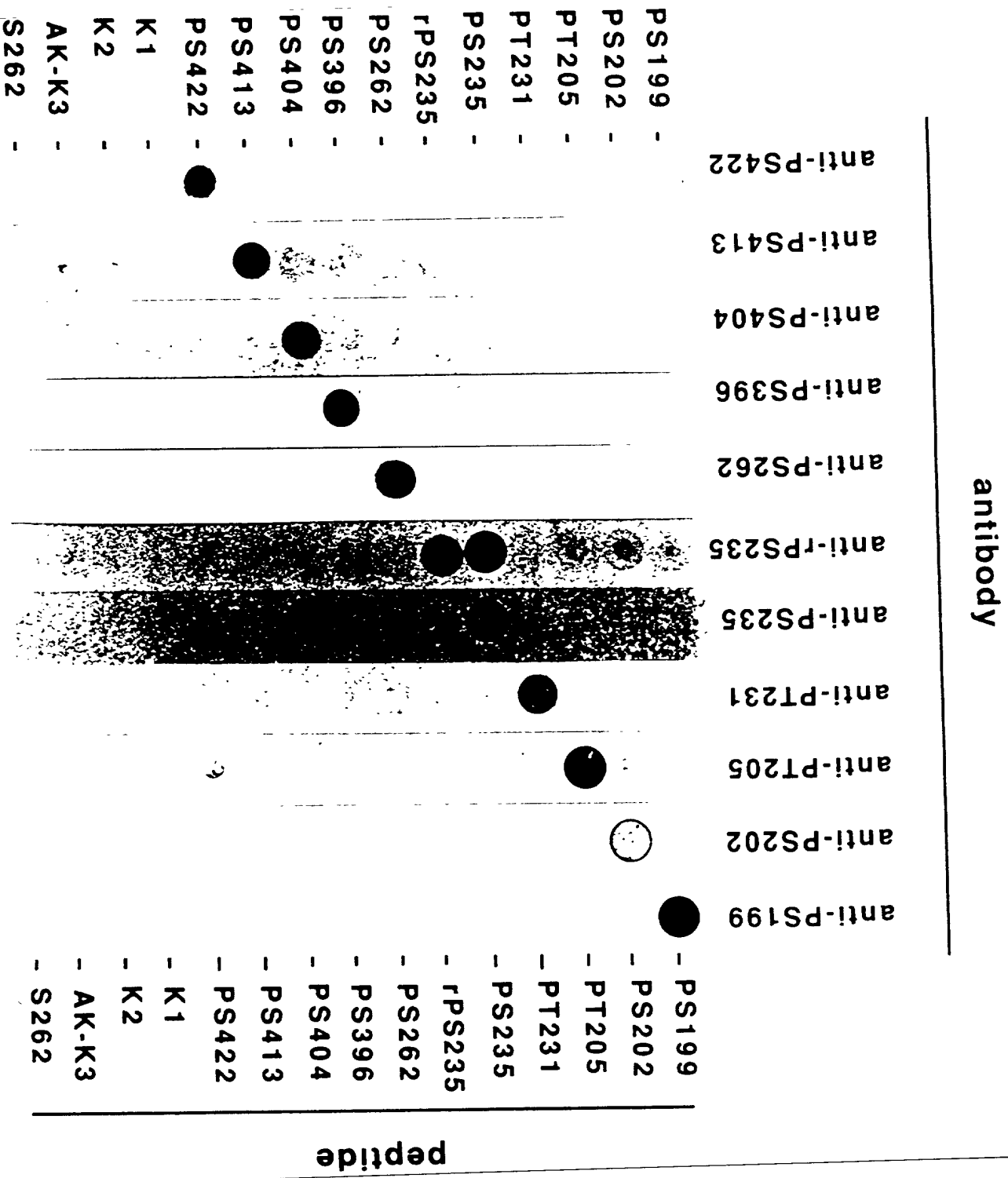


FIG. 2

TS

AD/N	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	26	27
A	A	A	A	A	N	N	N	N	N	N	N	N	N	A	A	A	A	A	A	A	A	A	A	A	A	A	
PHF	++	++	++	++	++	++	++	++	++	++	++	++	++	++	++	++	++	++	++	++	++	++	++	++	++	++	

PS 262
(S x 1500)

ID No. 1 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17 18 19 20 21 22 23 24 25 26 27

PS 413

(x 62.5)

ID No. 1 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17 18 19 20 21 22 23 24 25 26 27

PS 422

(S x 62.5)

ID No. 1 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17 18 19 20 21 22 23 24 25 26 27

Tau-C

(x 10000)

ID No. 1 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17 18 19 20 21 22 23 24 25 26 27

FIG. 3

SDSppt

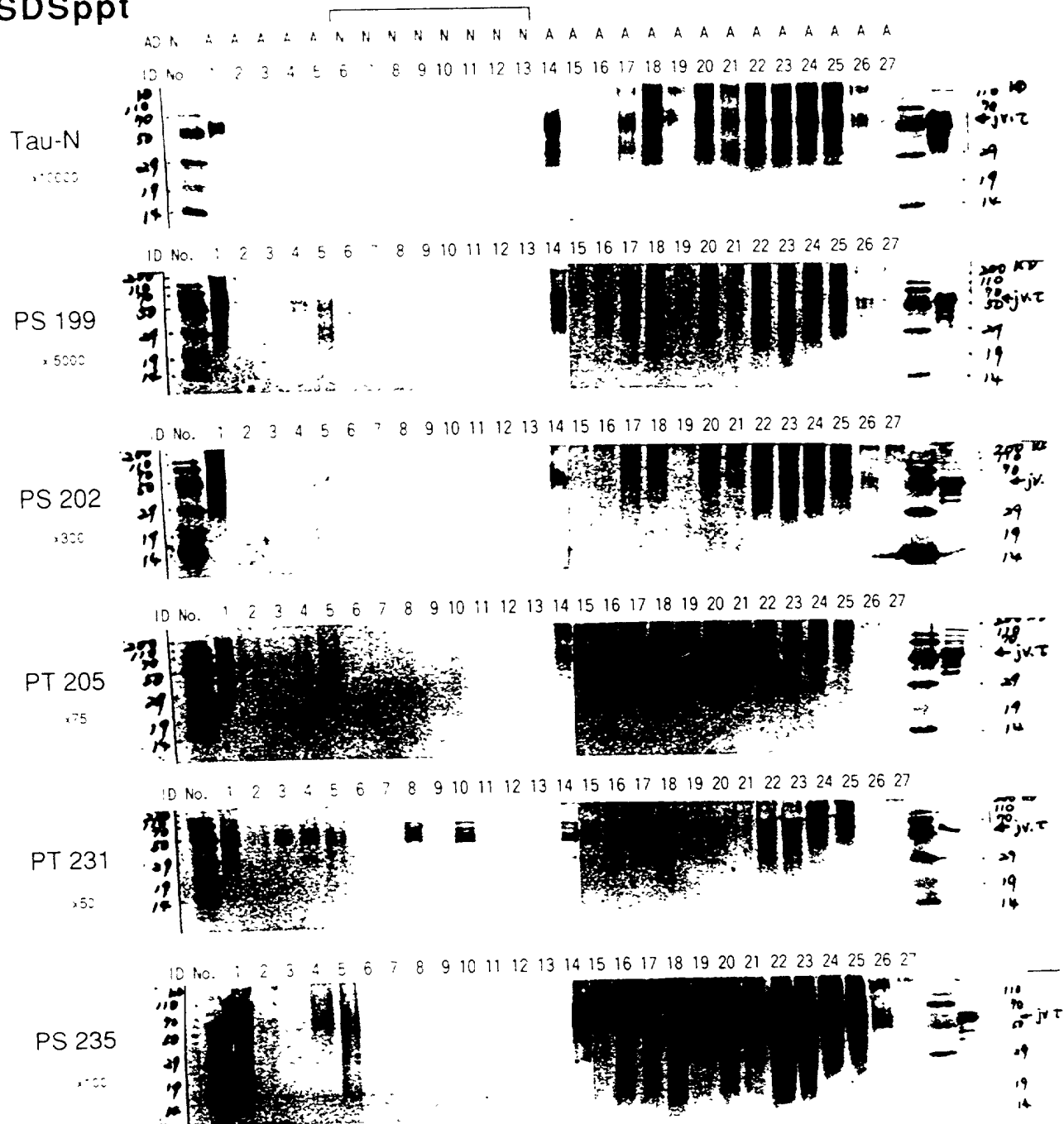


FIG. 4

SDSppt

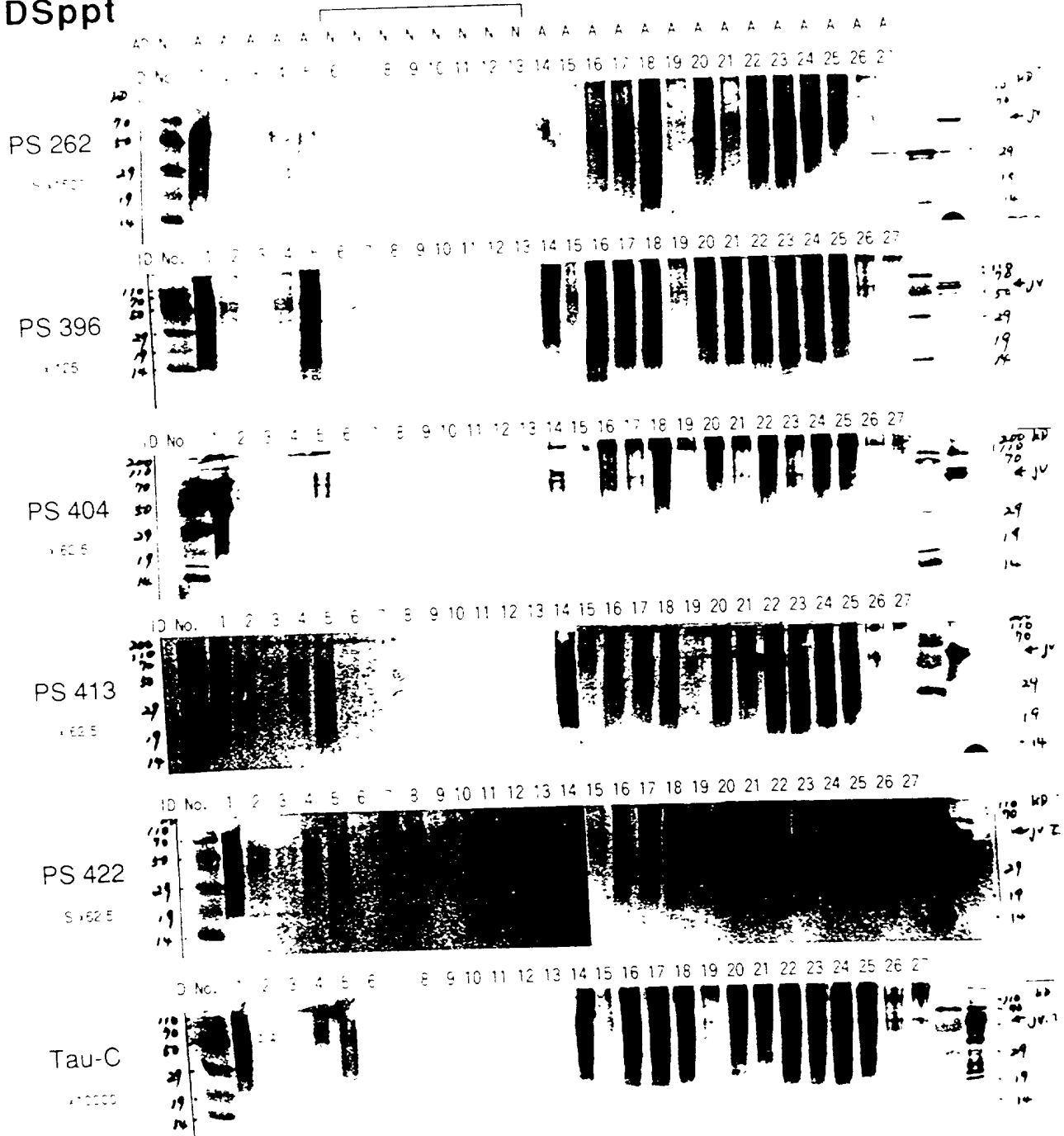
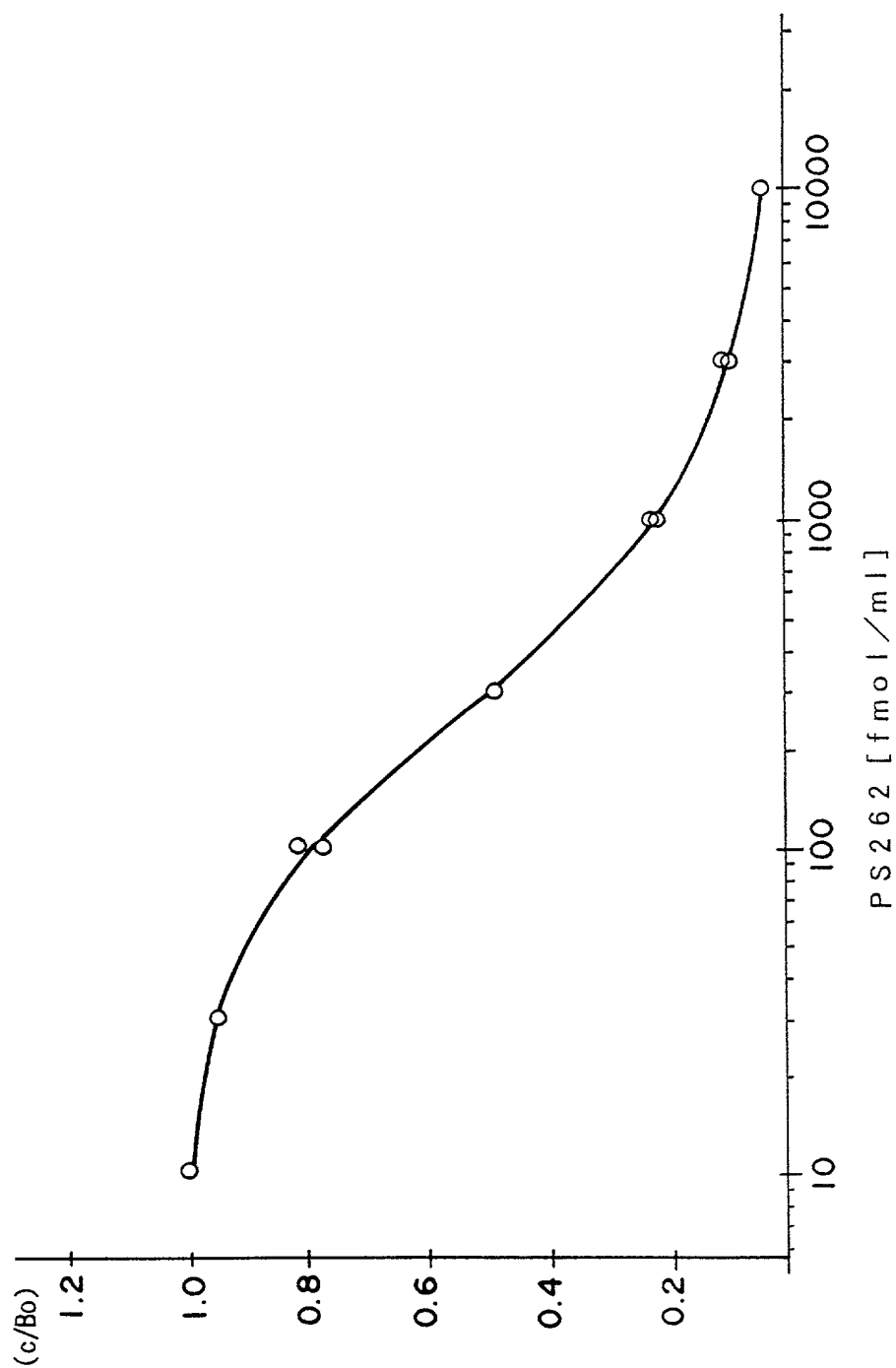
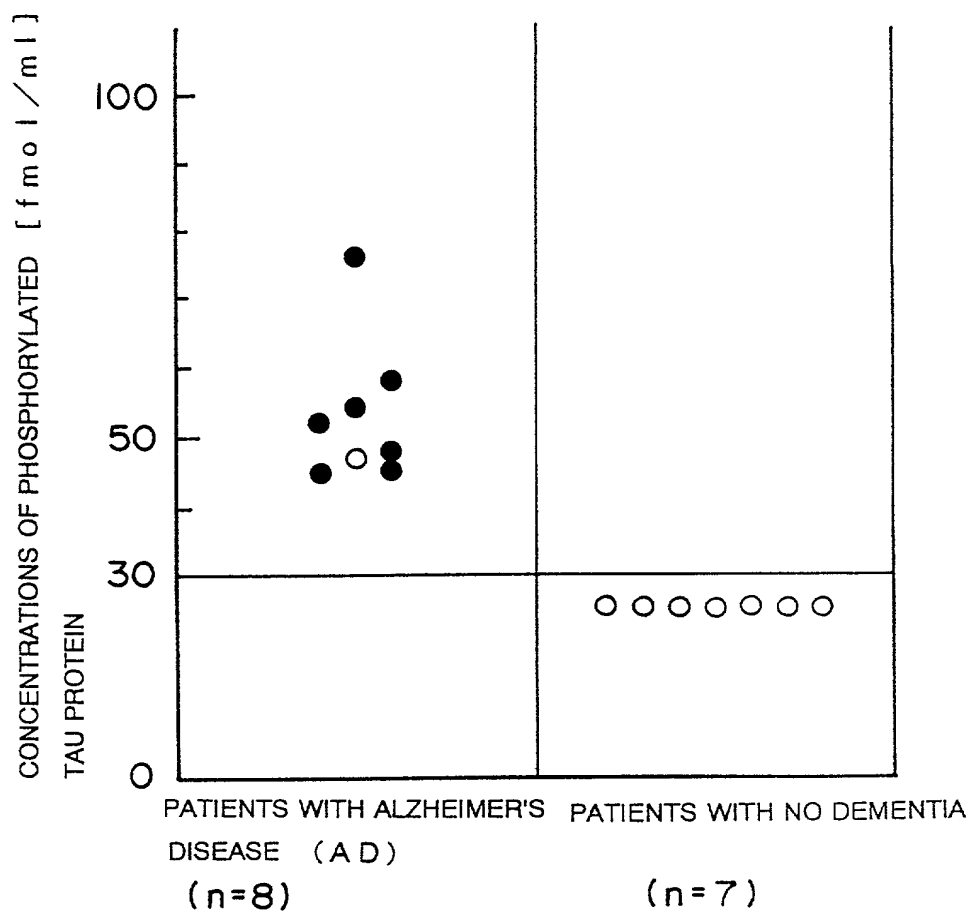


FIG. 5



[illegible]

DECLARATION AND POWER OF ATTORNEY FOR U.S. PATENT APPLICATION

() Original () Supplemental () Substitute () PCT () Design

As a below named inventor, I hereby declare that: my residence, post office address and citizenship are as stated below next to my name; that I verily believe that I am the original, first and sole inventor (if only one name is listed below) or an original, first and joint inventor (if plural inventors are named below) of the subject matter which is claimed and for which a patent is sought on the invention entitled:

Title: ANTI-PHOSPHORYLATED TAU PROTEIN ANTIBODIES AND METHOD FOR DETECTING ALZHEIMER'S DISEASE WITH THE USE OF THE SAME

of which is described and claimed in:

- (*) the attached specification, or
 () the specification in the application Serial No. _____ filed _____;
 and with amendments through _____ (if applicable), or
 (x) the specification in International Application No. PCT/ JP97/00804, filed March 13, 1997, and as amended
 on _____ (if applicable).

I hereby state that I have reviewed and understand the content of the above-identified specification, including the claims, as amended by any amendment(s) referred to above.

I acknowledge my duty to disclose to the Patent and Trademark Office all information known to me to be material to patentability as defined in Title 37, Code of Federal Regulations, §1.56.

I hereby claim priority benefits under Title 35, United States Code, §119 (and §172 if this application is for a Design) of any application(s) for patent or inventor's certificate listed below and have also identified below any application for patent or inventor's certificate having a filing date before that of the application on which priority is claimed:

COUNTRY	APPLICATION NO.	DATE OF FILING	PRIORITY CLAIMED
Japan	8-56090	March 13, 1996	YES

I hereby claim the benefit under Title 35, United States Code, §120 of any United States application(s) listed below and, insofar as the subject matter of each of the claims of this application is not disclosed in the prior United States application in the manner provided by the first paragraph of Title 35, United States Code, §112, I acknowledge the duty to disclose information material to patentability as defined in Title 37, Code of Federal Regulations, §1.56 which occurred between the filing date of the prior application and the national or PCT international filing date of this application.

APPLICATION SERIAL NO.	U.S. FILING DATE	STATUS: PATENTED, PENDING, ABANDONED

And I hereby appoint John T. Miller, Reg. No. 21,120; Michael R. Davis, Reg. No. 25,134; Matthew M. Jacob, Reg. No. 25,154; Jeffrey Noltan, Reg. No. 25,408; Warren M. Cheek, Jr., Reg. No. 33,367; Nils E. Pedersen, Reg. No. 33,145 and Charles R. Watts, Reg. No. 33,142, who together constitute the firm of WENDEROTH, LIND & PONACK, L.L.P., attorneys to prosecute this application and to transact all business in the U.S. Patent and Trademark Office connected therewith.

I hereby authorize the U.S. attorneys named herein to accept and follow instructions from Sera, Toyama, Matsukura, Kawaguchi & Koiwai as to any action to be taken in the U.S. Patent and Trademark Office regarding this application without direct communication between the U.S. attorneys and myself. In the event of a change in the persons from whom instructions may be taken, the U.S. attorneys named herein will be so notified by me.

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Residence & Citizenship	CITY	STATE OR COUNTRY	COUNTRY OF CITIZENSHIP
Post Office Address	ADDRESS	CITY	STATE OR COUNTRY ZIP CODE
Full Name of Seventh Inventor	FAMILY NAME	FIRST GIVEN NAME	SECOND GIVEN NAME
Residence & Citizenship	CITY	STATE OR COUNTRY	COUNTRY OF CITIZENSHIP
Post Office Address	ADDRESS	CITY	STATE OR COUNTRY ZIP CODE

I further declare that all statements made herein of my own knowledge are true, and that all statements on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code, and that such willful false statements may jeopardize the validity of the application or any patent issuing thereon.

1st Inventor Koichi Ishiguro Date April 9, 1999
 2nd Inventor Kazuki Sato Date April 9, 1999
 3rd Inventor Jun-Mi Park Date April 9, 1999
 4th Inventor Tsuneke Uchida Date April 9, 1999
 5th Inventor Kazutomo Imahori Date April 9, 1999
 6th Inventor _____ Date _____
 7th Inventor _____ Date _____

The above application may be more particularly identified as follows:

U.S. Application Serial No. _____ Filing Date _____
 Applicant Reference Number _____ Atty Docket No. _____
 Title of Invention _____